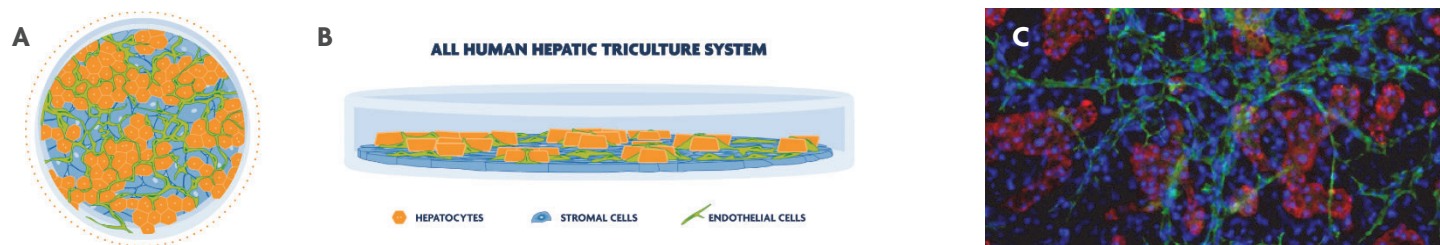


TruVIVO®

All-Human 2D+ Hepatic System

TruVivo is an all-human 2D+ hepatic system that combines primary human hepatocytes (PHHs) with primary human feeder cells (FCs) resulting in a human-relevant, reliable, and simplistic *in vitro* platform for compound testing.



A schematic representation of TruVivo showing (A) the top view of self-assembled hepatocyte colonies, (B) the side view of hepatocyte colonies integrated among the mixed feeder cell layer, and (C) a representative confocal laser microscopy image of TruVivo (blue: nuclei [DAPI], green: endothelial cells [CD31], red: hepatocytes [albumin]) revealing self-assembled hepatocyte colonies that mimic the microarchitecture of the human liver.

TruRELEVANCE

TruVivo is an **all-human 2D+ hepatic system that addresses the limitations of other traditional methods** by maintaining the morphological and functional properties of hepatocytes over several weeks. TruVivo can be established from a broad spectrum of donors, including prevalent disease states.

TruRELIABILITY

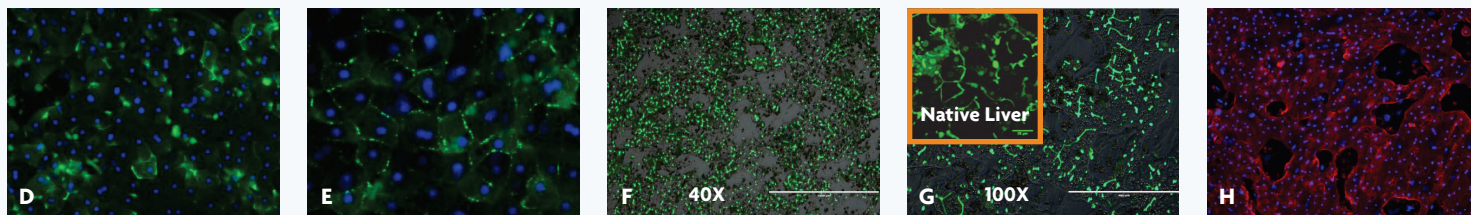
Hepatic Model	Limitation(s)	TruVivo Solution
Animal	Marked species differences versus human <i>in vivo</i>	All human cells; no species differences
Mono- and sandwich culture (MC and SC)	Short- to mid-term loss of hepatic morphology and functionality	Maintains hepatic morphology and functionality over prolonged periods up to 31 days
Isolated human hepatocytes in suspension (SHH)	Relatively short incubation time (≤ 6 h), limiting accurate evaluation of slowly metabolized compounds	Suitable to determine metabolic profiles and clearance of low-turnover compounds
Traditional Co-culture or Tri-Culture	Combine hepatocytes with one or more different types of non-human FCs	All-human FCs

TruVivo **is an easy-to-use platform that delivers equivalent or better performance** for predicting drug- and chemical-induced hepatotoxicity or metabolic clearance compared to other methods¹⁻¹⁰

TruRELIABILITY

1. Maintains viability and native hepatocellular morphology over several weeks:¹⁻⁴

- PHHs in TruVivo formed extensive anastomosing networks of bile canaliculi with both tight and gap junctions and can maintain hepatocyte morphology for 42 days *in vitro* with no collagen or Matrigel overlay (Figures D-H).¹⁻³
- Cell viability is maintained over time in TruVivo, as indicated by a lactase dehydrogenase (LDH) viability assay with no media change for up to 7 days.⁴



Tight Junction Marker: ZO-1

Gap Junction Marker: Cx-32

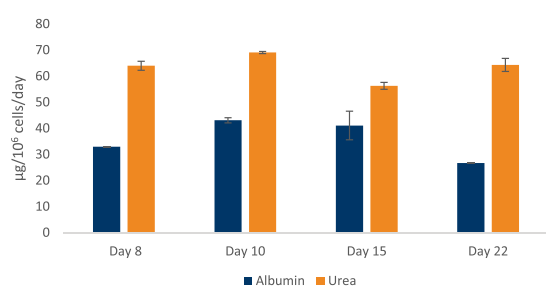
CDFDA fluorescent staining of bile canaliculi (Day 15)

Ck-18 (red)

2. Sustains hepatocellular function and metabolic activity over time.⁵

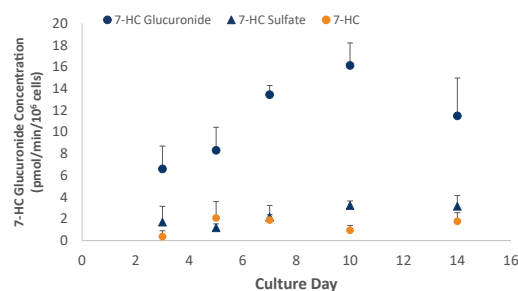
- Significantly higher albumin synthesis and urea production compared to PHHs in the SC monoculture over a 4-week culture period (Figure I).^{2,3}
- TruVivo generated Phase I and II metabolites more extensively than SHH (Figure J).⁶
- Hepatocytes in the 24- and 96-well formats maintained CYP and UGT activity, comparable to that of suspended hepatocytes, for as long as 31 days in culture (Figure K).^{1,7}

I Urea and Albumin Synthesis



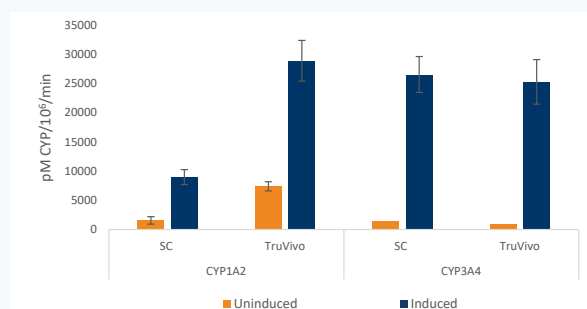
TruVivo is able to maintain albumin and urea levels in PHHs over a 22 day culture period.³

J Phase I and II Metabolic Pathway



Phase I and II metabolic pathways sustained in TruVivo over a 2-week period.⁶

K CYP Activity

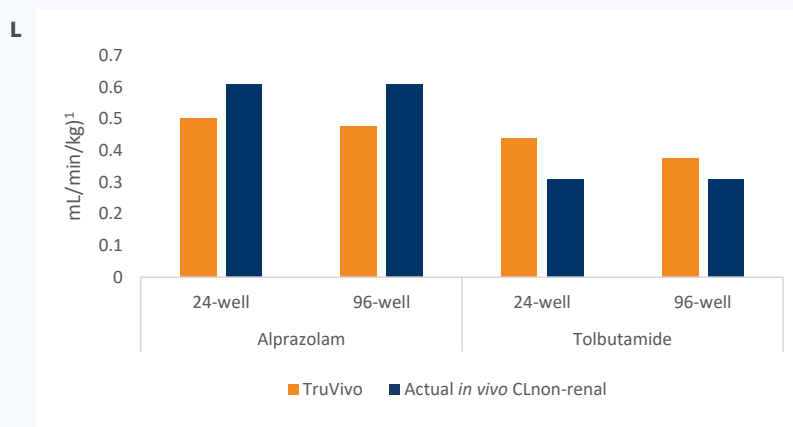


CYP1A2 and CYP3A4 induction in TruVivo compared to SC monoculture on day 4.¹

TruRELIABILITY

3. Suitable model for low-turnover compounds to predict *in vivo* metabolism:⁴

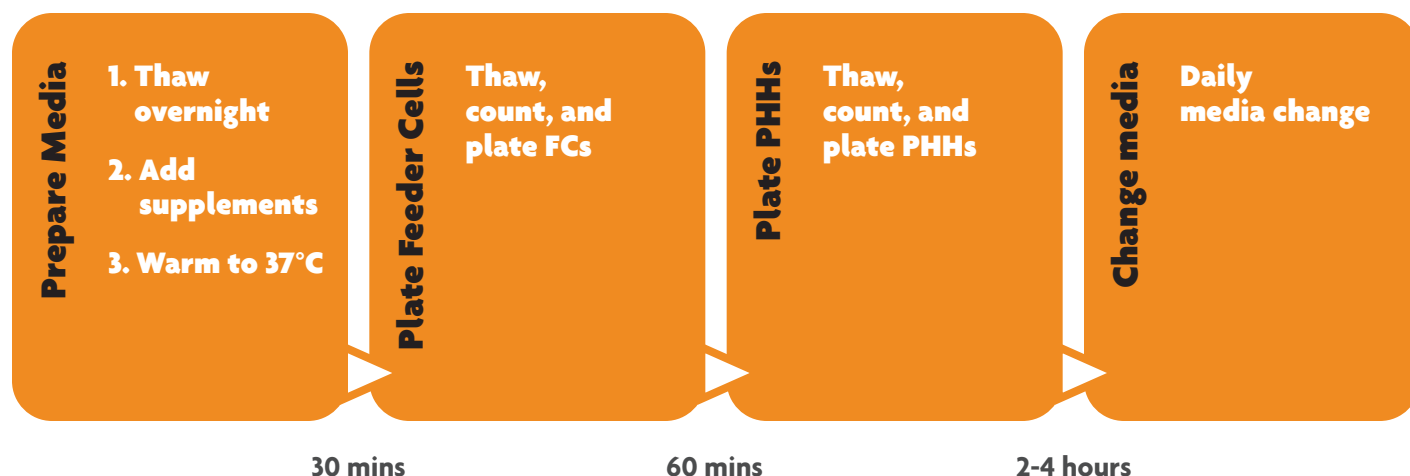
- TruVivo determined metabolic profiles, clearance of low-turnover compounds, and enzyme induction from healthy and diseased tissues in both the 24- and 96-well formats.^{4,6,8,9}
- TruVivo predicted hepatic clearance (CL) of low-turnover compounds within 2-fold of actual *in vivo* non-renal CL values, in both 24- and 96-well formats (Figure L).^{2,4}
- TruVivo evaluated thyroid hormone perturbation (key and associated events) after exposure to known nuclear receptor agonists.¹⁰



Comparison of predicted *in vitro* hepatic clearance values for alprazolam and tolbutamide to actual *in vivo* non-renal clearance.⁴

TruSIMPLICITY

The **2D+** system offers the simplicity and flexibility of a 2D model with the robustness of a 3D model. It is a convenient, stable, and reproducible platform that can be set up in a matter of hours in any standard laboratory equipped for cell culture and is suitable for a wide range of applications.⁸ This makes TruVivo a valuable tool for pharmacological and toxicological applications in drug development and risk assessment.



TruVIVO®

All-Human 2D+ Hepatic System

References

1. Weaver J, Odanga J, Wolf K, Stone T, Piekos S, Taub M, Thomas C, Byer-Alcorace A, Chen J, Lee JB, & LeCluyse E. Characterization of Morphology, Longevity and Functionality in an All-Human Cell-Based Triculture System. Data on file at LifeNet Health. Presented at SOT 2022.
2. Piekos S, Weaver J, Thomas C, Byer-Alcorace A, Odanga J, Wolf K, Chen J, Lee JB, LeCluyse E, & Taub M. Characterization of Clearance Mechanisms in an All-Human Cell-Based Triculture System. Presented at ISSX 2022.
3. Weaver J, Odanga J, Wolf K, Stone T, Piekos S, Taub M, Thomas C, Byer-Alcorace A, Chen J, Lee JB, & LeCluyse E. An All-Human Cell-Based Triculture System Distinguished Through Morphology, Longevity, and Functionality. Data on file at LifeNet Health. Presented at ICT 2022.
4. Byer-Alcorace A, Thomas C, Piekos S, Weaver J, Odanga J, Wolf K, Chen J, Lee JB, LeCluyse E, & Taub M. Utilizing a Long-Term All-Human Triculture System to Assess Hepatic Clearance of Low Turnover Drugs. Presented at GRC 2022.
5. Piekos S, Thomas C, Byer-Alcorace A, Weaver J, Odanga J, Wolf K, Chen J, Lee JB, LeCluyse E, & Taub M. Characterizing the Induction Potential of Major Cytochrome P450 Genes in a Novel All-Human Cell-Based Hepatocyte Triculture System. Presented at GRC 2022.
6. Maw H, Wang T, Raymond K, Byer-Alcorace A, Piekos S, Chan T, & Taub M. Assessing the In Vitro In Vivo Correlation of Small Molecule Metabolism in Three Long-Term Primary Human Hepatocyte Culture Models. Presented at ISSX 2022.
7. Thomas C, Byer-Alcorace A, Piekos S, Weaver J, Odanga J, Wolf K, Chen J, Lee JB, LeCluyse E, & Taub M. Evaluation of the Activity of Major Drug Metabolizing Enzymes in a Novel All-Human Hepatic Cell-Based Triculture System. Presented at GRC 2022.
8. Odanga J, Breathwaite E, Presnell S, LeCluyse E, & Weaver, J. Characterization of Primary Human Hepatocytes from Diseased and Healthy Livers in an All-human Cell-based Triculture System. Data on file at LifeNet Health. Presented at SOT 2023.
9. Murchison A, Wolf K, Breathwaite E, Weaver J, Treadwell M, Soldatow V, Chen J, LeCluyse E, & Lee JB. A Novel In Vitro All-Human Triculture Model That Maintains Structural Organization and Key Functions of Primary Hepatocytes Over Several Weeks. Data on file at LifeNet Health. Presented at ELRIG 2021.
10. Raza, A, Wolf K, Biven M, Stone T, Kellum S, LeCluyse E, LaRocca J, Settivari R, & Catalano S. In Vitro Triculture Hepatic Model to Evaluate Human Relevance of Chemical-Induced Thyroid Toxicity. Presented at SOT 2023.